

# Integrative Structure Validation Report ?

March 13, 2025 - 12:34 PM PDT

*The following software was used in the production of this report:*

*Integrative Modeling Validation Version 2.0*

*Python-IHM Version 1.8*

*MolProbity Version 4.5.2*

PDB ID	9A88
PDB-Dev ID	PDBDEV_00000373
Structure Title	Structure of the pre-incision complex in nucleotide excision repair
Structure Authors	Yu, J.; Yan, C.Y.; Paul, T.; Brewer, L.; Tsutakawa, S.E.; Tsai, C.-L.; Hamdan, S.; Tainer, J.A.; Ivanov, I.
Deposited on	2024-03-11

*This is a PDB-IHM IM Structure Validation Report for a publicly released PDB-IHM entry.*

*We welcome your comments at [helpdesk@pdb-ihm.org](mailto:helpdesk@pdb-ihm.org)*

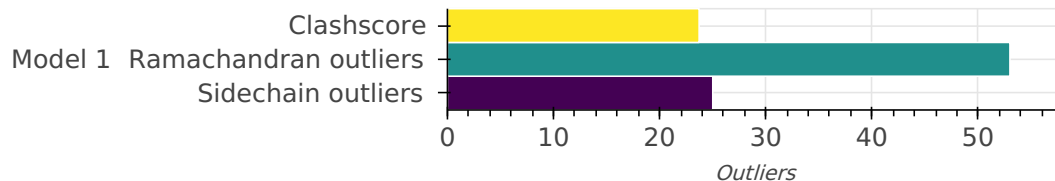
*A user guide is available at [https://pdb-ihm.org/validation\\_help.html](https://pdb-ihm.org/validation_help.html) with specific help available everywhere you see the ? symbol.*

*List of references used to build this report is available [here](#).*

## Overall quality ?

*This validation report contains model quality assessments for all structures, data quality and fit to model assessments for SAS and crosslinking-MS datasets. Data quality and fit to model assessments for other datasets and model uncertainty are under development. Number of plots is limited to 256.*

Model Quality: MolProbity Analysis



### Ensemble information ?

*This entry consists of 0 distinct ensemble(s).*

### Summary ?

*This entry consists of 1 model(s). A total of 25 datasets were used to build this entry.*

### Representation ?

*This entry has 1 representation(s).*

ID	Model(s)	Entity ID	Molecule name	Chain(s) [auth]	Total residues	Rigid segments	Flexible segments	Model coverage/ Starting model coverage (%)	Scale
1	1	1	General transcription and DNA repair factor IIH helicase subunit XPB	A	720	-	34-203, 248-720	89.31 / 100.00	Atomic
		2	General transcription and DNA repair factor IIH helicase subunit XPD	B	760	-	1-760	100.00 / 0.00	Atomic
		3	General transcription factor IIH subunit 4	C	441	-	1-441	100.00 / 100.00	Atomic

ID	Model(s)	Entity ID	Molecule name	Chain(s) [auth]	Total residues	Rigid segments	Flexible segments	Model coverage/ Starting model coverage (%)	Scale
		4	General transcription factor IIH subunit 2	D	377	-	1-377	100.00 / 100.00	Atomic
		5	General transcription factor IIH subunit 3	E	292	-	1-292	100.00 / 0.00	Atomic
		6	General transcription factor IIH subunit 5	F	66	-	1-66	100.00 / 100.00	Atomic
		7	DNA repair protein complementing XP-A cells	G	273	-	1-273	100.00 / 0.00	Atomic
		8	General transcription factor IIH subunit 1	H	154	-	1-154	100.00 / 100.00	Atomic
		9	DNA excision repair protein ERCC-5	I	985	-	1-296, 733-985	55.74 / 100.00	Atomic
		10	DNA repair endonuclease XPF Gene: ERCC4, ERCC11, XPF	J	227	-	1-227	100.00 / 100.00	Atomic
		11	DNA excision repair	K	198	-	1-198	100.00 / 100.00	Atomic
		12	Replication protein A 70 kDa DNA-binding subunit, N-terminally processed	L	434	-	1-434	100.00 / 100.00	Atomic
		13	Replication protein A 14 kDa subunit	M	115	-	1-115	100.00 / 100.00	Atomic

ID	Model(s)	Entity ID	Molecule name	Chain(s) [auth]	Total residues	Rigid segments	Flexible segments	Model coverage/ Starting model coverage (%)	Scale
		14	Replication protein A 32 kDa subunit	N	225	-	1-225	100.00 / 100.00	Atomic
		15	DNA (66-MER)	O [X]	66	-	1-66	100.00 / 100.00	Atomic
		16	DNA (66-MER)	P [Y]	66	-	1-66	100.00 / 100.00	Atomic
		17	IRON/SULFUR CLUSTER	Q [B]	Non-polymeric	-	-	Not available / Not available	Atomic
		18	ZINC ION	R [D]	Non-polymeric	-	-	Not available / Not available	Atomic
				S [D]					
				T [D]					
				U [E]					
				V [E]					
				W [G]					
				Z [L]					
		19	MAGNESIUM ION	X [I]	Non-polymeric	-	-	Not available / Not available	Atomic
				Y [J]					
		20	water	AA [Y]	Non-polymeric	-	-	Not available / Not available	Atomic

### Datasets used for modeling ?

There are 25 unique datasets used to build the models in this entry.

ID	Dataset type	Database name	Data access code
4	Crosslinking-MS data	Not available	<a href="#">10.1038/s41467-019-10745-5</a>
1	Experimental model	PDB	<a href="#">6RO4</a>
2	3DEM volume	EMDB	<a href="#">EMD-4970</a>

ID	Dataset type	Database name	Data access code
3	Experimental model	PDB	<a href="#">6TUW</a>
5	De Novo model	Not available	Not available
6	De Novo model	Not available	Not available
7	De Novo model	Not available	Not available
8	Experimental model	PDB	<a href="#">6SXA</a>
9	Experimental model	PDB	<a href="#">6SXB</a>
10	Experimental model	PDB	<a href="#">2BGW</a>
11	De Novo model	Not available	Not available
12	Experimental model	PDB	<a href="#">4GOP</a>
13	Experimental model	PDB	<a href="#">6I52</a>
14	Experimental model	PDB	<a href="#">1JMC</a>
15	Experimental model	PDB	<a href="#">1L1O</a>
16	Experimental model	PDB	<a href="#">2JNW</a>
17	Experimental model	PDB	<a href="#">4MQV</a>
18	De Novo model	Not available	Not available
19	De Novo model	Not available	Not available
20	De Novo model	Not available	Not available
21	De Novo model	Not available	Not available
22	De Novo model	Not available	Not available
23	De Novo model	Not available	Not available
24	De Novo model	Not available	Not available
25	De Novo model	MODEL ARCHIVE	<a href="#">ma-2chon</a>

## Methodology and software

*This entry is a result of 1 distinct protocol(s).*

Step number	Protocol ID	Method name	Method type	Method description	Number of computed models	Multi state modeling	Multi scale modeling
				To construct a model of the pre-incision complex (PlnC), we systematically examined the cryo-EM structures and densities of human apo-TFIIH, TFIIH/XPA/DNA, and			

Step number	Protocol ID	Method name	Method type	Method description	Number of computed models	Multi state modeling	Multi scale modeling
				<p>XPF/ERCC1, the NMR structure of XPA-ERCC1, and the X-ray structures of the XPG catalytic core and RPA-ssDNA (RPA70, RPA32, and RPA14). The TFIIH/XPA/DNA structure (PDB ID: 6RO4 and EMDB accession code: EMD-4970) was the starting point for model building. The PlnC hybrid model has an NER bubble size of 23 nucleotides, matching the 27-nucleotide optimal length of the excision products and the XPF and XPG incision patterns.</p> <p>FEN1 shares 30% sequence identity with the XPG catalytic core (PDB ID: 6TUR, 6TUW, and 6VBH). Thus, we modeled DNA-bound XPG based on the human FEN1/DNA X-ray structure (PDB ID: 5UM9). XPG positioning into the hybrid model was based on existing XL-MS data. In addition, positioning of the XPG core required placement of the 3' DNA junction 8 nucleotides away from the expected position of the DNA lesion near XPD's His135 residue.</p> <p>The two XPG gateway helices (GH1, residues 82-126) and (GH2, residues 734-761) were predicted with AlphaFold2 and positioned in the gap between XPD's Arch and Fe-S domains in accordance with the crosslink data. The XPD-anchor domain (residues 157-296) was predicted by AlphaFold2 and fitted into the TFIIH/XPA/DNA cryo-EM</p>			

Step number	Protocol ID	Method name	Method type	Method description	Number of computed models	Multi state modeling	Multi scale modeling
1	1	None	None	<p>density. The loop connecting GH1 and the XPD anchor was built with Modeller. To model XPF/ERCC1, we used the cryo-EM structures of XPF/ERCC1 (PDB ID: 6SXA and 6SXB). We first docked the XPF nuclease domain to the 5' junction. The catalytic metal was oriented 3A away from the scissile phosphodiester bond. Mg<sup>2+</sup> ion coordination was based on the Aeropyrum pernix SNF2 structure (PDB ID: 2BGW). A water molecule was placed between Mg<sup>2+</sup> ion and the DNA backbone phosphate group. The ERCC1 (HhH)<sub>2</sub> domain was oriented to interact with the ssDNA through two DNA hairpins based on the 6SXB structure. The long linkers from the ERCC1 central domain to the ERCC1 (HhH)<sub>2</sub> (residues 214-230) and from the XPF nuclease domain to the XPF (HhH)<sub>2</sub> (residues 817-847) were built with Modeller. The SF2 helicase-like N-terminal domain of XPF was omitted from the hybrid PlnC model due to lack of sufficient structural or biochemical restraints. To model RPA, we used following X-ray structures: Ustilago maydis RPA/ssDNA (PDB ID: 4GOP), yeast RPA/ssDNA (PDB ID: 6I52) and human RPA (PDB ID: 1JMC and 1L1O). The RPA70AB/ssDNA complex was modeled by</p>	None	False	False

Step number	Protocol ID	Method name	Method type	Method description	Number of computed models	Multi state modeling	Multi scale modeling
				<p>superimposing the yeast RPA/ssDNA structure (PDB ID: 1JMC) onto the human apo-RPA 70AB (PDB ID: 6I52). Within PInC, only RPA70A, 70B, and 70C can engage DNA due to the size of the NER bubble. RPA70AB was placed close to the 3' junction where it interacts with XPG. We reoriented RPA70C to bind ssDNA near the 5' junction. The RPA70C/ssDNA was modeled by aligning the <i>Ustilago maydis</i> RPA/ssDNA structure (PDB ID: 4GOP) with the human trimer core structure (PDB ID: 1L1O). The orientation of RPA32D and RPA14 follows from the placement of the RPA70C module as they are all connected, forming the trimer core (70C/32D/14). To model XPA, we used the following structures: the cryo-EM TFIIH/XPA/DNA structure (PDB ID: 6RO4), the NMR structure of XPA/ERCC1 (PDB ID: 2JNW), and the human X-ray structure of RPA32C/Smarca11 N-terminus (PDB ID: 4MQV). The XPA N-terminal extension (residues 1-103), which includes the RPA32C binding helix (residues 22-40), and the C-terminal extension (beta-domain) (residues 235-273) lacked known structural homologues and were modeled using AlphaFold2. The beta-domain was fitted into the TFIIH/XPA/DNA density. To position XPA's N-terminal</p>			



Step number	Protocol ID	Method name	Method type	Method description	Number of computed models	Multi state modeling	Multi scale modeling
				helix (residues 22-40) we used the X-ray structure of KPA52C/Smad4 L N terminus. To assemble the complete PInC model, we also modelled loop regions of TFIIH's core subunits (XPB, XPD, p44, p34, and p52) into the TFIIH/XPA/DNA density.			

There are 6 software packages reported in this entry.

ID	Software name	Software version	Software classification	Software location
1	<a href="https://alphafold.ebi.ac.uk/">AlphaFold2</a>	Not available	model building	<a href="https://alphafold.ebi.ac.uk/">https://alphafold.ebi.ac.uk/</a>
2	<a href="https://salilab.org/modeller/">Modeller</a>	10.40	model building	<a href="https://salilab.org/modeller/">https://salilab.org/modeller/</a>
3	<a href="https://www.ebi.ac.uk/jdispatcher/msa/clustalo">Clustal Omega</a>	Not available	sequence alignments	<a href="https://www.ebi.ac.uk/jdispatcher/msa/clustalo">https://www.ebi.ac.uk/jdispatcher/msa/clustalo</a>
4	<a href="https://www2.mrc-lmb.cam.ac.uk/personal/pemsley/coot/">Coot</a>	0.9.8.92	real-space refinement	<a href="https://www2.mrc-lmb.cam.ac.uk/personal/pemsley/coot/">https://www2.mrc-lmb.cam.ac.uk/personal/pemsley/coot/</a>
5	<a href="https://phenix-online.org/">Phenix</a>	1.20.1	real-space refinement	<a href="https://phenix-online.org/">https://phenix-online.org/</a>
6	<a href="https://www.cgl.ucsf.edu/chimera/">UCSF Chimera</a>	1.18	model visualization	<a href="https://www.cgl.ucsf.edu/chimera/">https://www.cgl.ucsf.edu/chimera/</a>

## Data quality ?

### Crosslinking-MS

At the moment, data validation is only available for crosslinking-MS data deposited as a fully *compliant* dataset in the [PRIDE Crosslinking](#) database. Correspondence between crosslinking-MS and entry entities is established using *pyHMMER*. Only residue pairs that passed the reported threshold are used for the analysis. The values in the report have to be interpreted in the context of the experiment (i.e. only a minor fraction of in-situ or in-vivo dataset can be used for modeling).

Crosslinking-MS dataset is not available in the [PRIDE Crosslinking](#) database.

### 3DEM volume

Validation for this section is under development.

## Model quality ?

For models with atomic structures, MolProbity analysis is performed. For models with coarse-grained or multi-scale structures, excluded volume analysis is performed.

### Standard geometry: bond outliers ?

*There are 24 bond length outliers in this entry (0.06% of 41860 assessed bonds). A summary is provided below.*

Chain	Res	Type	Atoms	Z	Observed (Å)	Ideal (Å)	Model ID (Worst)	Models (Total)
B	191	PRO	CG-CD	25.40	0.64	1.50	1	1
J	9	MET	SD-CE	11.34	1.51	1.79	1	1
L	192	PRO	CB-CG	10.98	0.94	1.49	1	1
B	191	PRO	N-CD	10.48	1.62	1.47	1	1
I	273	LEU	CG-CD1	10.19	1.18	1.52	1	1
J	10	ARG	CG-CD	7.66	1.29	1.52	1	1
E	147	MET	SD-CE	6.48	1.63	1.79	1	1
H	120	ARG	CG-CD	6.43	1.33	1.52	1	1
B	266	LEU	CG-CD1	6.39	1.31	1.52	1	1
B	191	PRO	N-CA	6.39	1.37	1.47	1	1
A	380	MET	CG-SD	6.20	1.65	1.80	1	1
B	191	PRO	CB-CG	5.99	1.79	1.49	1	1
H	128	VAL	CB-CG1	5.89	1.33	1.52	1	1
A	570	GLU	CG-CD	5.86	1.37	1.52	1	1
H	116	GLU	CB-CG	5.74	1.35	1.52	1	1
B	386	LEU	CG-CD2	5.59	1.34	1.52	1	1
E	42	MET	SD-CE	5.47	1.65	1.79	1	1
L	192	PRO	N-CD	5.25	1.40	1.47	1	1
J	188	MET	SD-CE	5.17	1.66	1.79	1	1
L	192	PRO	N-CA	4.80	1.54	1.47	1	1
D	47	MET	SD-CE	4.72	1.67	1.79	1	1
G	257	TYR	CD1-CE1	4.35	1.25	1.38	1	1
K	135	ARG	CB-CG	4.22	1.39	1.52	1	1
G	257	TYR	CE2-CZ	4.21	1.28	1.38	1	1

### Standard geometry: angle outliers ?

*There are 42 bond angle outliers in this entry (0.07% of 57190 assessed bonds). A summary is provided below.*

Chain	Res	Type	Atoms	Z	Observed (Å)	Ideal (Å)	Model ID (Worst)	Models (Total)
L	192	PRO	CB-CG-CD	23.09	179.97	106.10	1	1
B	191	PRO	N-CD-CG	21.99	70.21	103.20	1	1
L	192	PRO	CA-CB-CG	20.80	64.97	104.50	1	1
L	192	PRO	N-CD-CG	19.65	73.73	103.20	1	1
B	191	PRO	CA-N-CD	14.69	91.43	112.00	1	1
L	192	PRO	CA-N-CD	11.99	95.21	112.00	1	1
B	191	PRO	CA-CB-CG	11.21	83.20	104.50	1	1
E	147	MET	CG-SD-CE	9.90	79.13	100.90	1	1
A	511	TRP	C-N-CA	9.84	139.41	121.70	1	1
J	10	ARG	CG-CD-NE	9.43	132.75	112.00	1	1
H	120	ARG	CG-CD-NE	8.76	131.28	112.00	1	1
B	190	CYS	CA-C-N	8.65	129.87	116.90	1	1
B	191	PRO	N-CA-CB	7.30	94.97	103.00	1	1
B	215	PRO	CA-N-CD	6.96	102.25	112.00	1	1
A	570	GLU	CG-CD-OE1	6.39	103.70	118.40	1	1
P	29	DA	O3'-P-OP1	6.01	89.97	108.00	1	1
H	116	GLU	CG-CD-OE2	5.66	131.42	118.40	1	1
G	168	LYS	C-N-CA	5.60	131.78	121.70	1	1
H	20	TYR	C-N-CA	5.50	131.59	121.70	1	1
B	190	CYS	CA-C-O	5.48	111.48	120.80	1	1
H	116	GLU	CB-CG-CD	5.47	121.89	112.60	1	1
K	170	GLU	CA-CB-CG	5.39	103.32	114.10	1	1
B	427	THR	C-N-CA	5.33	131.30	121.70	1	1
J	11	GLU	CG-CD-OE1	5.27	106.28	118.40	1	1
J	158	THR	C-N-CA	5.23	131.12	121.70	1	1
B	424	ARG	C-N-CA	5.19	112.36	121.70	1	1
A	643	VAL	C-N-CA	4.91	112.87	121.70	1	1
J	148	THR	C-N-CA	4.82	130.37	121.70	1	1
G	62	VAL	C-N-CA	4.80	130.35	121.70	1	1
J	122	LEU	C-N-CA	4.70	130.16	121.70	1	1
B	278	GLU	N-CA-C	4.66	124.05	111.00	1	1
B	369	ARG	C-N-CA	4.43	129.68	121.70	1	1
K	135	ARG	CA-CB-CG	4.41	122.93	114.10	1	1
H	120	ARG	CD-NE-CZ	4.33	130.46	124.40	1	1
L	258	THR	C-N-CA	4.27	129.38	121.70	1	1

Chain	Res	Type	Atoms	Z	Observed (Å)	Ideal (Å)	Model ID (Worst)	Models (Total)
B	191	PRO	CB-CG-CD	4.22	92.59	106.10	1	1
G	90	VAL	C-N-CA	4.16	129.18	121.70	1	1
B	370	LYS	CA-CB-CG	4.15	122.41	114.10	1	1
B	257	ASP	C-N-CA	4.15	114.23	121.70	1	1
G	127	ASP	C-N-CA	4.10	114.33	121.70	1	1
L	249	LYS	N-CA-C	4.09	122.44	111.00	1	1
B	190	CYS	O-C-N	4.05	116.53	123.00	1	1

### Too-close contacts ?

The following all-atom clashscore is based on a MolProbity analysis. All-atom clashscore is defined as the number of clashes found per 1000 atoms (including hydrogen atoms). The table below contains clashscores for all atomic models in this entry.

Model ID	Clash score	Number of clashes
1	23.71	1903

There are 1903 clashes. The table below contains the detailed list of all clashes based on a MolProbity analysis. Bad clashes are  $\geq 0.4$  Angstrom. The output is limited to 100 rows.

Atom 1	Atom 2	Clash(Å)	Model ID (Worst)	Models (Total)
B:501:GLN:HB2	J:188:MET:HE2	1.14	1	1
I:273:LEU:HD13	K:48:ASN:HA	1.13	1	1
H:116:GLU:HG2	H:120:ARG:HD2	1.11	1	1
G:249:GLU:HB2	G:257:TYR:CE1	1.08	1	1
B:410:TYR:HB3	B:414:PHE:HD2	1.08	1	1
G:18:ALA:HB2	M:2:MET:HE2	1.06	1	1
B:353:SER:HB2	L:273:GLN:HG2	1.05	1	1
C:140:GLU:OE2	C:144:HIS:HD2	1.00	1	1
L:252:GLY:O	L:273:GLN:NE2	0.99	1	1
B:424:ARG:O	I:288:HIS:CE1	0.99	1	1
C:211:MET:HE2	C:240:SER:HB2	0.98	1	1
J:157:GLU:HA	J:160:PRO:HG2	0.97	1	1
G:237:ARG:HG3	G:239:THR:H	0.95	1	1
E:143:TYR:CZ	E:147:MET:HE1	0.94	1	1
H:116:GLU:HG3	H:120:ARG:HH11	0.93	1	1
G:249:GLU:HB2	G:257:TYR:HE1	0.93	1	1
B:257:ASP:O	B:260:GLN:HG3	0.93	1	1
B:410:TYR:HB3	B:414:PHE:CD2	0.93	1	1
B:543:GLN:H	P:37:DG:H5"	0.92	1	1

Atom 1	Atom 2	Clash(Å)	Model ID (Worst)	Models (Total)
G:167:LYS:HZ1	L:404:ARG:HH22	0.92	1	1
D:56:ASP:HB3	D:61:MET:HE2	0.92	1	1
K:149:THR:HG21	K:182:LYS:HD3	0.91	1	1
H:116:GLU:CG	H:120:ARG:HD2	0.91	1	1
C:140:GLU:OE2	C:144:HIS:CD2	0.90	1	1
B:336:LEU:HD21	B:358:LEU:HD12	0.89	1	1
B:422:ASP:H	B:427:THR:HG22	0.89	1	1
B:501:GLN:OE1	K:194:PHE:HB3	0.88	1	1
A:568:LEU:HD11	A:606:PHE:HB3	0.88	1	1
H:116:GLU:CG	H:120:ARG:HH11	0.88	1	1
C:132:ASP:O	C:136:GLU:HG2	0.88	1	1
G:113:MET:HG2	G:114:ASP:H	0.88	1	1
A:584:THR:HA	K:185:ARG:HD3	0.87	1	1
H:116:GLU:HG3	H:120:ARG:NH1	0.86	1	1
A:570:GLU:OE1	K:170:GLU:HG2	0.86	1	1
B:273:ILE:HD12	B:276:THR:HB	0.86	1	1
I:288:HIS:CE1	I:289:TYR:O	0.86	1	1
K:136:VAL:HG13	K:154:LEU:HD12	0.85	1	1
J:35:GLY:O	J:150:LEU:HD12	0.85	1	1
D:246:GLN:HE21	E:251:TYR:HB3	0.84	1	1
G:167:LYS:NZ	L:404:ARG:HH22	0.84	1	1
B:657:MET:HE3	B:689:LYS:HB3	0.84	1	1
N:37:ILE:HB	N:80:GLU:H	0.84	1	1
G:168:LYS:HB2	G:169:ASN:HA	0.84	1	1
B:503:ALA:O	J:185:ARG:NH1	0.84	1	1
B:280:ARG:HH12	B:387:GLU:HG2	0.84	1	1
J:9:MET:HE3	K:135:ARG:HB3	0.84	1	1
J:151:ALA:HB1	J:153:THR:H	0.83	1	1
J:69:MET:HG2	J:76:PRO:HB3	0.82	1	1
I:273:LEU:HD13	K:48:ASN:CA	0.82	1	1
D:64:GLN:N	D:64:GLN:OE1	0.82	1	1
I:271:GLY:HA2	K:51:PRO:HD3	0.82	1	1
B:288:LEU:HD13	B:324:ARG:HB3	0.82	1	1
B:360:GLY:H	L:253:VAL:H	0.81	1	1
N:36:ILE:HG12	N:80:GLU:HG2	0.81	1	1

Atom 1	Atom 2	Clash(Å)	Model ID (Worst)	Models (Total)
B:302:ASP:HB2	B:380:ARG:HE	0.81	1	1
B:176:ASN:OD1	B:177:LEU:N	0.81	1	1
O:3:DT:O2	P:65:DG:N2	0.81	1	1
B:292:LEU:HD22	B:374:PHE:CD1	0.81	1	1
B:414:PHE:HE1	B:437:CYS:HB2	0.81	1	1
A:360:ARG:HH12	A:433:GLN:HB3	0.80	1	1
B:136:SER:HB3	B:389:THR:HB	0.80	1	1
B:346:VAL:HG23	L:220:ASN:HD21	0.80	1	1
G:67:LYS:HB2	K:9:ARG:HD2	0.80	1	1
I:198:PRO:HB2	I:254:MET:SD	0.80	1	1
I:169:MET:HA	I:169:MET:HE2	0.80	1	1
C:81:THR:OG1	C:89:GLN:NE2	0.80	1	1
B:424:ARG:HE	B:426:PRO:HG2	0.79	1	1
I:39:LEU:HD12	P:53:DT:H2'	0.79	1	1
D:263:MET:HE2	D:297:ILE:HG13	0.79	1	1
G:76:ILE:HG21	K:45:ARG:HB3	0.79	1	1
B:497:ARG:NH2	B:707:ASN:OD1	0.79	1	1
D:263:MET:CE	D:297:ILE:HG13	0.79	1	1
B:543:GLN:HA	B:546:GLU:HB2	0.79	1	1
J:36:ASP:HB2	J:69:MET:HE1	0.79	1	1
B:426:PRO:HA	I:290:ILE:HD11	0.78	1	1
B:138:THR:HB	B:155:CYS:HB3	0.78	1	1
C:211:MET:HE2	C:240:SER:CB	0.78	1	1
H:16:GLU:HG3	H:17:MET:HE2	0.78	1	1
A:569:LYS:HG3	K:184:ARG:HH21	0.78	1	1
N:109:MET:HA	N:109:MET:HE2	0.78	1	1
O:39:DG:H2''	O:40:DC:H5''	0.77	1	1
B:359:SER:HB2	L:253:VAL:C	0.77	1	1
B:284:GLU:OE2	B:378:ARG:NH2	0.77	1	1
A:380:MET:SD	A:381:TRP:CD1	0.76	1	1
L:253:VAL:HG13	L:278:ASP:HB2	0.76	1	1
B:416:ILE:HG22	B:435:PHE:HD1	0.76	1	1
G:32:ARG:HG2	M:83:VAL:HB	0.76	1	1
G:228:ARG:HE	K:176:PRO:HD2	0.76	1	1
B:292:LEU:HD22	B:374:PHE:HD1	0.76	1	1

Atom 1	Atom 2	Clash(Å)	Model ID (Worst)	Models (Total)
I:913:LYS:HA	P:66:DT:H2"	0.76	1	1
A:577:LYS:HD3	A:604:THR:HG21	0.75	1	1
B:354:PRO:HA	B:415:THR:HA	0.75	1	1
G:91:VAL:HG11	G:138:LEU:H	0.75	1	1
J:155:ASP:O	K:148:LYS:NZ	0.75	1	1
D:340:GLY:HA3	E:146:ARG:HH12	0.75	1	1
B:386:LEU:HA	I:103:ARG:HE	0.75	1	1
H:63:ILE:HG23	H:119:ARG:HH21	0.75	1	1
G:18:ALA:HA	M:2:MET:H	0.75	1	1
B:424:ARG:HH22	L:228:LYS:HZ2	0.75	1	1
E:6:ASP:HB2	E:155:GLN:HB2	0.74	1	1

### Torsion angles: Protein backbone ?

*In the following table, Ramachandran outliers are listed. The Analysed column shows the number of residues for which the backbone conformation was analysed.*

Model ID	Analysed	Favored	Allowed	Outliers
1	4722	4111	558	53

*There are 53 unique backbone outliers. Detailed list of outliers are tabulated below.*

Chain	Res	Type	Models (Total)
A	71	HIS	1
A	110	PRO	1
A	145	LYS	1
A	150	ASP	1
A	199	LEU	1
B	354	PRO	1
B	355	PRO	1
B	426	PRO	1
B	428	ILE	1
C	32	PRO	1
D	16	GLU	1
D	36	VAL	1
D	37	PHE	1
D	68	PRO	1
D	264	ALA	1
E	82	PHE	1

Chain	Res	Type	Models (Total)
E	92	ASN	1
G	8	LEU	1
G	13	ALA	1
G	18	ALA	1
G	20	LEU	1
G	21	PRO	1
G	43	LEU	1
G	61	ASN	1
G	91	VAL	1
G	97	VAL	1
G	118	MET	1
G	128	ASN	1
G	129	CYS	1
G	170	PRO	1
G	237	ARG	1
G	240	ILE	1
G	241	VAL	1
H	55	ILE	1
I	913	LYS	1
J	10	ARG	1
J	149	ALA	1
J	151	ALA	1
J	152	ILE	1
J	153	THR	1
J	155	ASP	1
J	159	LEU	1
L	190	ARG	1
L	218	ILE	1
L	237	GLY	1
L	243	VAL	1
L	245	ILE	1
L	251	GLY	1
L	259	ASN	1
L	270	ASN	1
L	312	GLN	1



Chain	Res	Type	Models (Total)
N	43	ALA	1
N	131	SER	1

### Torsion angles : Protein sidechains ?

*In the following table, sidechain rotameric outliers are listed. The Analysed column shows the number of residues for which the sidechain conformation was analysed.*

Model ID	Analysed	Favored	Allowed	Outliers
1	4194	3176	993	25

*There are 25 unique sidechain outliers. Detailed list of outliers are tabulated below.*

Chain	Res	Type	Models (Total)
A	38	VAL	1
A	65	MET	1
A	187	HIS	1
A	398	ASP	1
B	42	MET	1
B	136	SER	1
B	257	ASP	1
B	266	LEU	1
B	444	ILE	1
B	501	GLN	1
B	546	GLU	1
B	708	LEU	1
C	41	ARG	1
C	359	MET	1
C	424	MET	1
D	295	CYS	1
D	315	LEU	1
G	48	TYR	1
G	59	MET	1
G	100	PHE	1
H	30	SER	1
I	247	ILE	1
J	69	MET	1
J	123	TRP	1
L	299	CYS	1

## Fit of model to data used for modeling ?

### Fit of model(s) to crosslinking-MS data

#### Restraint types

Restraint types are summarized in the table below. Restraints assigned "*by-residue*" are interpreted as between CA atoms. Restraints between coarse-grained beads are indicated as "*coarse-grained*".

*Restraint group* represents a set of crosslinking restraints applied collectively in the modeling.

There are 21 crosslinking restraints combined in 21 restraint groups.

Linker	Residue 1	Atom 1	Residue 2	Atom 2	Restraint type	Distance, Å	Count
BS3	ALA	None	GLY	None	upper bound	30.0	1
BS3	ALA	None	LEU	None	upper bound	30.0	2
BS3	ASP	None	GLU	None	upper bound	30.0	2
BS3	GLU	None	LEU	None	upper bound	30.0	4
BS3	ASP	None	PHE	None	upper bound	30.0	1
BS3	GLY	None	PHE	None	upper bound	30.0	1
BS3	ARG	None	PHE	None	upper bound	30.0	1
BS3	LEU	None	PHE	None	upper bound	30.0	1
BS3	ARG	None	GLU	None	upper bound	30.0	1
BS3	LEU	None	SER	None	upper bound	30.0	1
BS3	ASP	None	GLN	None	upper bound	30.0	1
BS3	ASP	None	LEU	None	upper bound	30.0	1
BS3	LEU	None	LEU	None	upper bound	30.0	2
BS3	ARG	None	ASP	None	upper bound	30.0	1
BS3	GLN	None	LEU	None	upper bound	30.0	1

#### Distograms of individual restraints

Restraints with identical thresholds are grouped into one plot. Only the best distance per restraint per model group/ensemble is plotted. Inter- and intramolecular (including self-links) restraints are also grouped into one plot. Distance for a restraint between coarse-grained beads is calculated as a minimal distance between shells; if beads intersect, the distance will be reported as 0.0. A bead with the highest available resolution for a given residue is used for the assessment.

#### Satisfaction of restraints

Satisfaction of restraints is calculated on a *restraint group* (a set of crosslinking restraints applied collectively in the modeling) level. Satisfaction of a restraint group depends on satisfaction of individual restraints in the group and the conditionality (all/any). A restraint group is considered satisfied, if the condition was met in at least one model of the model group/ensemble. The number of measured restraints can be smaller than the total number of restraint groups if crosslinks involve non-modeled residues. Only deposited models are used for validation right now.

State group	State	Model group	# of Deposited models/Total	Restraint group type	Satisfied (%)	Violated (%)	Count (Total=21)
1	1	1	1/1	All	Not available	Not available	0

#### Per-model satisfaction rates in ensembles

*Every point represents one model in a model group/ensemble. Where possible, boxplots with quartile marks are also plotted.*

#### 3DEM volume

Validation for this section is under development.

#### Fit of model to data used for validation ?

Validation for this section is under development.

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